

MINIREVIEW

Impact of Culture-Independent Studies on the Emerging Phylogenetic View of Bacterial Diversity

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INTRODUCTION

Our perspective on microbial diversity has improved enormously over the past few decades. In large part this has been due to molecular phylogenetic studies that objectively relate organisms. Phylogenetic trees based on gene sequences are maps with which to articulate the elusive concept of biodiversity. Thus, comparative analyses of small-subunit rRNA (16S or 18S rRNA) and other gene sequences show that life falls into three primary domains, *Bacteria*, *Eucarya*, and *Archaea* (51, 52). Based on rRNA trees, the main extent of Earth's biodiversity is microbial. Our knowledge of the extent and character of microbial diversity has been limited, however, by reliance on the study of cultivated microorganisms. It is estimated that >99% of microorganisms observable in nature typically are not cultivated by using standard techniques (1).

Recombinant DNA and molecular phylogenetic methods have recently provided means for identifying the types of organisms that occur in microbial communities without the need for cultivation (see references 1, 20, and 35 for reviews). Results from application of these methods to a number of diverse environments confirm that our view of microbial diversity was limited and point to a wealth of novel and environmentally important diversity yet to be studied (34). It is the aim of this review to collate, compare, and incorporate the results of the environmental sequence-based studies into the context of known bacterial diversity. We discuss the sequence data at the taxonomic level of the phylogenetic division because divisions constitute first-order clades for describing the breadth of bacterial diversity. Although we have yet to determine even the outlines of the bacterial tree, common threads are beginning to emerge that revise our current views of bacterial diversity and distribution in the environment.

PHYLOGENETIC DIVERSITY IN THE BACTERIAL DOMAIN

In 1987, Woese described the bacterial domain as comprised of about 12 natural relatedness groups, based mainly on analyses of familiar cultivated organisms such as cyanobacteria, spirochetes, and gram-positive bacteria (all of which, based on rRNA sequence divergence, display greater evolutionary depth than plants, animals, and fungi) (51). These relatedness groups have variously been called "kingdoms," "phyla," and "divi-

sions"; we use the latter term. For the purposes of this review we define a bacterial division purely on phylogenetic grounds as a lineage consisting of two or more 16S rRNA sequences that are reproducibly monophyletic and unaffiliated with all other division-level relatedness groups that constitute the bacterial domain. We judge reproducibility by the use of multiple tree-building algorithms, bootstrap analysis, and varying the composition and size of data sets used for phylogenetic analyses. The typical interdivisional rRNA sequence difference is 20 to 25%. For comparison, the 16S rRNAs of *Escherichia coli* and *Pseudomonas aeruginosa*, both representatives of the γ group of *Proteobacteria*, differ overall by about 15%; the 16S rRNAs of *E. coli* and *Bacillus subtilis* ("low-G + C gram-positive bacterial" division) differ by about 23%.

At the current stage in the phylogenetic classification of *Bacteria*, divisions are not consistently named or taxonomically ranked. rRNA-defined divisions are identified by classes (e.g., *Proteobacteria* [41] and *Actinobacteria* [42]), orders (e.g., *Thermotogales* and *Aquificales*), families (e.g., *Chlorobiaceae*), generic names such as the *Nitrospira* group (11), or common names such as the green nonsulfur (GNS) bacteria and low-G + C gram-positive bacteria (51). Division-level nomenclature has not even been consistent between studies, so some divisions are identified by more than one name. For instance, green sulfur bacteria is synonymous with *Chlorobiaceae*; high-G + C gram-positive bacteria is synonymous with *Actinobacteria* and *Actinomycetales*. Indeed, it probably is premature to standardize taxonomic rankings for bacterial divisions at this point when our picture of microbial diversity is likely still incomplete and the topology of the bacterial tree is still unresolved.

In the past decade the number of identifiable bacterial divisions has more than tripled to about 40 due in significant part to culture-independent phylogenetic surveys of environmental microbial communities (21, 34). These analyses rely on sequences of rRNA genes obtained by cloning directly from environmental DNA or, as in the majority of studies, after amplification by the PCR (1, 20, 35). Figure 1 represents the division-level diversity of the bacterial domain as inferred from representatives of the approximately 8,000 bacterial 16S rRNA gene sequences currently available. Although 36 divisions are shown in Fig. 1, several other division-level lineages are indicated by single environmental sequences (9, 21, 37), suggesting that the number of bacterial divisions may be well over 40. Several of the described divisions are well represented by cultivated strains and were the first to be characterized phylogenetically (51). The majority of the bacterial divisions, however, are poorly represented by cultured organisms. Indeed, 13 of the 36 divisions shown in Fig. 1 are characterized only by environmental sequences (shown outlined) and are termed "candidate divisions" to indicate their unstantiated status as

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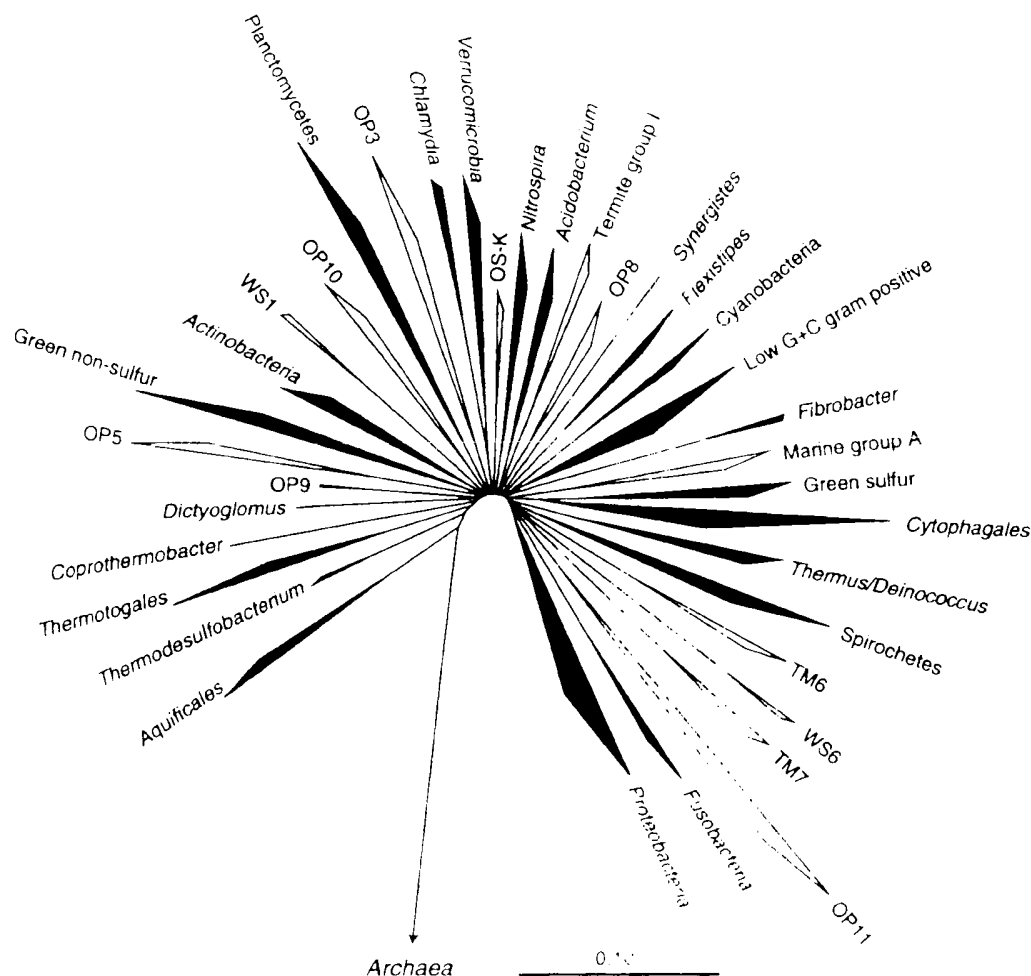


FIG. 1. Evolutionary distance tree of the bacterial domain showing currently recognized divisions and putative (candidate) divisions. The tree was constructed using the ARB software package (with the Lane mask and Olsen rate-corrected neighbor joining algorithm) and a reference database modified from the March 1997 ARB database release (43). Division-level groupings of two or more sequences are depicted as wedges. The length of the wedge reflects the branching depth of the representatives selected for a particular division. Divisions which have cultivated representatives are shown in black; divisions represented only by environmental sequences are shown in outline. The scale bar indicates 0.1 change per nucleotide. The aligned 16S rRNA sequences used for this figure and Fig. 3 through 6 are available from <http://cab2.berkeley.edu/pacelab/176.htm>.

new bacterial divisions (21). One of these candidate divisions, OP11, is now sufficiently well represented by environmental sequences to conclude that it constitutes a major bacterial group (see below). Phylogenetic studies so far have not resolved branching orders of the divisions; bacterial diversity is seen as a fan-like radiation of division-level groups (Fig. 1). The exception to this, however, is the *Aquificales* division, which branches most deeply in the bacterial tree in most analyses.

BACTERIAL DIVERSITY AND DISTRIBUTION IN THE ENVIRONMENT

Culture dependent studies indicate that representatives of some bacterial divisions are cosmopolitan in the environment, whereas others appear restricted to certain habitats (39). Culture independent studies so far conducted reflect and expand this view. Table 1 summarizes the environmental distribution of sequences by habitat type, compiled from most of the available 16S rRNA-based clonal analyses: 86 studies contributing nearly 3,000 sequences. An expanded version of this table that

details division-level representation in the individual studies is available at <http://cab2.berkeley.edu/pacelab/176.htm>. Table 1 includes only divisions for which representatives have been detected in at least two independent studies and for which at least one near-complete 16S rRNA gene sequence is known. Table 1 is, therefore, not an exhaustive listing of potential division-level diversity for all studies.

Sequence representatives of several bacterial divisions have been identified in a wide range of habitats, suggesting the cosmopolitan or ubiquitous distribution of the corresponding organisms in the environment and, potentially, their broad metabolic capabilities. Some of these cosmopolitan divisions are well-known from cultivation studies; however, others are little known or have not yet been detected by cultivation. Figure 2 summarizes the representation of selected cosmopolitan divisions by sequences of cultivated and uncultivated organisms. The *Proteobacteria* (purple photosynthetic bacteria and related groups), *Cytophaga* (*Bacteroides*-*Cytophaga*-*Flexibacter* group), and the two gram-positive divisions, *Actinobacteria* and low-G+C gram-positive bacteria, are well represented by cultivation

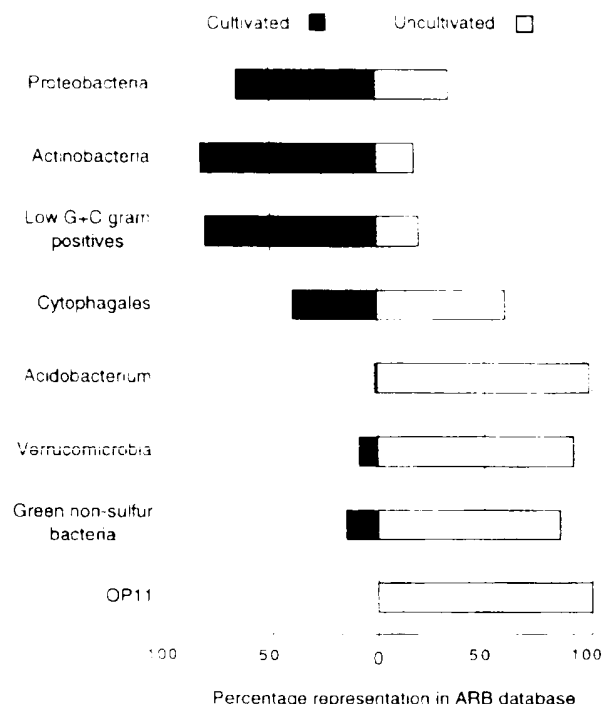


FIG. 2. Relative representation in selected cosmopolitan bacterial divisions of 16S rRNA sequences from cultivated and uncultivated organisms. Results were compiled from 5,224 and 2,918 sequences from cultivated and uncultivated organisms, respectively.

ed organisms and therefore are familiar to us in principle. These four divisions account for 90% of all cultivated bacteria characterized by 16S rRNA sequences and approximately 70% of the environmental sequences collated in Table 1. By contrast, other cosmopolitan divisions revealed by clonal analyses, such as *Acidobacterium*, *Verrucomicrobia*, GNS bacteria, and OP11, are poorly represented by sequences from cultivated organisms (Fig. 2) and consequently are little known with regard to their general properties. Although many of the bacterial divisions occur widely, others seem to occupy a more limited range of habitats (Table 1). All cultivated representatives of *Aquificales*, for instance, are thermophilic hydrogen metabolizers, and all environmental sequences of *Aquificales* have been obtained only from high-temperature environments. This suggests a specialized habitat niche for this group. Alternatively, the apparently limited environmental distribution may simply reflect a sampling or methodological artifact and representatives of such divisions may be present in a wider range of habitats, but not yet detected.

The database of environmental rRNA sequences is compromised in resolving some phylogenetic issues by a large number of relatively short sequences. More than half of the sequences collated in Table 1 are less than 500 nucleotides (nt) long, which represents only one-third of the total length of 16S rRNA. This is due to an unfortunate trend in many environmental studies of sequencing only a portion of the gene in the belief that a few hundred bases of sequence data is sufficient for phylogenetic purposes. Indeed, 500 nt is sufficient for placement if some longer sequence is closely related (>90% identity in homologous nucleotides) to the query sequence. In the case of novel sequences, >85% identical to known sequences, however, >500 nt is usually insufficient comparative information to place the sequence accurately in a phylogenetic tree and can even be misleading.

TABLE 1. Summary of 16S rRNA-based clonal analyses of diversity of uncultivated bacteria^a

Habitat type	No. of studies	No. of sequences ^a	<i>Proteobacteria</i> ^b						Bacterial division ^c
			α	β	γ	δ	ϵ		
Geothermal	10	212	•						Cytophagales
Soil	16	743	•					•	Actinobacteria
Marine	23	687	•		•			•	Low-G + C gram positives ^d
Freshwater	4	107	•						Acidobacterium
Wastewater	4	430	•		•				Verrucomicrobia
Pollutant associated	7	202	•						Spirochete ^e
Acid metal leaching	2	21			•				Nitrospira
Subsurface	6	229		•					GNS
Symbionts and commensals	10	280			•				OP11
Disease associated ^f	3	7							Planctomycetes
Totals	86	2,918							Green sulfur
									FM7
									FM6
									<i>Thermus</i> / <i>Durum</i> <i>Coccus</i>
									Cyanobacteria
									<i>Synechococcus</i>
									OPS
									Fe mate group I
									OSK
									<i>Chlamydia</i>
									OP3
									OP11
									WS1
									OPS
									Marine group A
									<i>Exiguobacter</i>
									<i>Flavobacter</i>
									<i>Deinococcus</i>
									<i>Thermotoga</i>
									<i>Thermoplasma</i>
									<i>Thiomargarita</i>
									<i>Thiobacillus</i>

^a An expanded version of this table detailing individual studies is available at <http://crab2.berkeley.edu/pocetib176.htm>.

^b Incidence of division-level representatives in studies of particular habitat types ranked from most represented to least represented divisions.

^c Excluding organelles.

^d *Proteobacteria* are presented at the subdivision level due to the extensive sequence representation of this division.

^e Cannot establish as a monophyletic group in all analyses.

Since all but 4 (40, 46, 49, 50) of the 86 studies collated in Table 1 were conducted using PCR to amplify rDNA from extracted environmental DNA, the question arises as to whether molecular analyses accurately reflect the division-level diversity that occurs in the environment. It is well established that PCR-associated artifacts such as differential amplification of different rDNA templates (36, 44), sensitivity to rRNA gene copy number (12), PCR primer specificity (48), sensitivity to template concentration (6), amplification of contaminant rDNA (45), and formation of chimeric sequences (23) may skew our assessment of microbial diversity. Most of the studies collated in Table 1, however, analyzed tens to hundreds of clones, so it seems likely that these studies have sampled the main types of sequences in the communities examined. We believe, acknowledging the caveats of the methodology, that the clonal analyses collated in Table 1 probably include the most abundant (metabolically active) bacterial sequence types in the samples analyzed, likely representing the members of the communities that are involved in the principal metabolic activities, such as carbon cycling.

ABUNDANT BUT LITTLE-KNOWN BACTERIAL DIVISIONS

The rRNA sequence studies of environmental organisms probably identify the abundant organisms in the environments studied and, therefore, account for the organisms that participate significantly in the maintenance of the communities. Because of their abundance in the environment, representatives of some poorly studied phylogenetic divisions are predicted to play significant roles in environmental chemistry. Examples of such divisions, which because of their potential environmental significance merit study, are the *Acidobacterium* division, the *Verrucomicrobia*, the GNS bacteria, and candidate division OP11.

***Acidobacterium* division.** The *Acidobacterium* group is a newly recognized bacterial division with only three cultivated representatives: *Acidobacterium capsulatum* (18), *Holophaga foetida* (26), and *Geothrix fermentans* (28). Figure 3 is a phylogenetic dendrogram of this group, including selected environmental representatives. The limited physiological information known about these organisms provides few clues to properties that might be general throughout the division. *Acidobacterium* is a moderately acidophilic aerobic heterotroph; *Holophaga* and *Geothrix* are strict anaerobes that ferment aromatic compounds and acetate, respectively. The majority of sequences that make up this division, however, are from environmental clones. At least eight monophyletic subdivisions in the *Acidobacterium* group are identified by phylogenetic analyses (Fig. 3 [24, 29]). We define a subdivision as a lineage comprised of two or more 16S rRNA sequences within a division that are reproducibly monophyletic and unaffiliated with all other representatives of that division. *Acidobacterium* subdivisions 1, 3, 4, and 6 are well represented by environmental clone sequences from independent studies, yet no cultivated strains are known with the exception of subdivision 1, represented by *A. capsulatum*. The widespread occurrence of environmental sequences belonging to the *Acidobacterium* division (Table 1) suggests that members of this group are ecologically significant constituents of many ecosystems, particularly soil communities. They have been detected in every clonal analysis of soils (with a wide range of chemical properties), as well as in other habitats, including a peat bog, acid mine drainage, a contaminated aquifer, a hot spring, a freshwater lake, and a sample of the Atlantic ocean from a depth of 1,000 m (Fig. 3). In situ single cell analyses with fluorescent hybridization probes specific for *Acidobacterium* subdivision 6 small subunit

rRNA indicate that this subdivision is morphologically diverse (29), as expected for a broad phylogenetic group. Members likely are metabolically diverse as well: the depth of phylogenetic diversity (depth of branching) in the *Acidobacterium* division is nearly as great as in the *Proteobacteria*.

***Verrucomicrobia*.** *Verrucomicrobia* is a newly proposed division of *Bacteria* (17) represented by a handful of isolates: *Verrucomicrobium spinosum* (after which the division is named) (47), four *Prostheco bacter* species (17), and three strains of ultramicrobacteria (22). *Verrucomicrobia* and *Prostheco bacter* are prosthecae bacteria isolated from freshwater, and the ultramicrobacteria, "dwarf-cell" strains only about 0.1 μm^3 in volume, were isolated from a soil habitat. All of these isolates preferentially use sugars as growth substrates. Culture-independent analyses indicate that the *Verrucomicrobia*, like members of the *Acidobacterium* division, are widespread in the environment and abundant, particularly in soils (Table 1). Figure 4 shows a dendrogram of representatives of the *Verrucomicrobia*. Several monophyletic subdivisions are seen, only two of which are represented by the cultivated strains. Clone sequences of this division from soil are predominantly from members of the phylogenetically broad subdivisions 2 and 3. The abundance of these two groups suggests their ecological importance. For instance, the abundance of one representative of *Verrucomicrobia* subdivision 2 (EA25) was estimated by PCR at 10^7 to 10^8 cells per g of a pasture soil sample, 1 to 10% of the total microbial content (25).

In our phylogenetic analyses we consistently find that the division *Chlamydia* is a specific sister group of the *Verrucomicrobia*. We find no support for the notion (17, 30, 47) of a specific relatedness of the planctomycetes with the *Verrucomicrobia*.

GNS bacteria. The GNS bacteria have been recognized as a division-level bacterial group for over a decade (51). Even today, however, this division is still represented by only a few isolates. The cultured representatives have a wide range of phenotypes, from anoxygenic photosynthesis (*Chloroflexus*) to thermophilic organotrophy (*Thermomicrobium*). Figure 5 shows the relatedness groups of GNS bacteria detected in the environment. It is apparent from the dendrogram that all of the cultivated representatives except the chlorinated hydrocarbon-reducing *Dehalococcoides ethenogenes* (31) are related in subdivision 3, together with several clone sequences from a hot spring, a rice paddy, and activated sludge (data not shown). By contrast, most of the environmental sequences described to date fall into a different relatedness group, subdivision 1, with no cultivated representatives. Considering the wide variety of habitats that have contributed GNS sequences (Fig. 5, Table 1), particularly to GNS subdivision 1, members of this division likely play significant roles in the environment.

Candidate division OP11. Candidate division OP11 is a recently proposed novel bacterial division for which there is no reported cultivated representative (19, 21). However, several independent clonal studies have reported environmental sequences that together form the OP11 clade. Figure 6 shows a dendrogram of the known environmental sequence representatives of the division, with five subdivisions currently identifiable. OP11 sequences all have highly atypical sequence signatures for the domain *Bacteria* (51), and they have low sequence identities, only about 80%, to sequences outside the OP11 division. This may be due to higher-than-average mutation rates in OP11 rRNAs, as has been suggested for other groups such as the planctomycetes (27). OP11 sequences have been obtained from a variety of habitats including several different types of soil, freshwater sediments, the deep subsurface, and hot springs (Table 1), suggesting that members of the division

Acidobacterium division

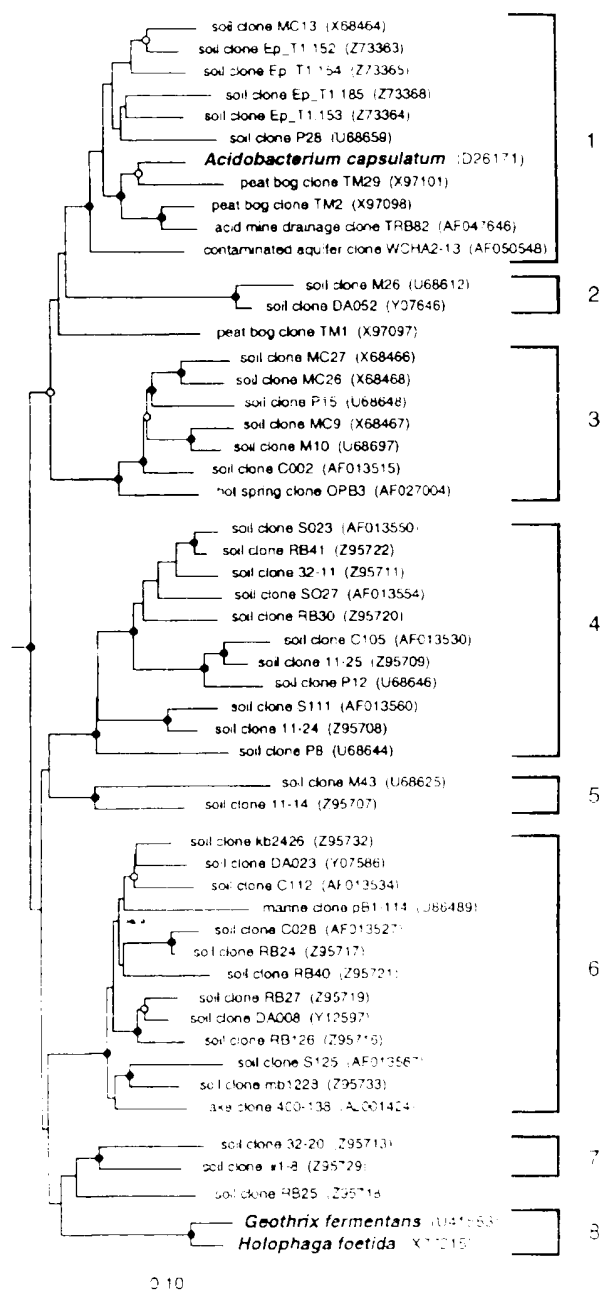


FIG. 3. Phylogenetic dendrogram of the *Acidobacterium* division. Names of cultivated organisms are shown in bold. The habitat source of each environmental sequence is indicated before the clone name. GenBank accession numbers are listed parenthetically. Subdivisions of the tree were as described for Fig. 1. The robustness of the topology presented was estimated by bootstrap resampling of independent distance, parsimony, and rate-corrected maximum-likelihood analyses as previously described (2). Distance and parsimony analyses were conducted using test version 4.4b1 of PAUP*, written by D. M. L. Swafford. Branch points supported (bootstrap values of >75%) by most or all of the three analyses are indicated by filled circles; open circles indicate branch points not fully supported (bootstrap values of 50 to 74% by most or all of the three analyses); without circles are not resolved. Bootstrap values of 100% are indicated by open circles. The scale bar indicates 0.1 change per nucleotide.

play significant ecological roles. Until cultivated representatives of the OP11 division are characterized, little beyond the general properties of *Bacteria* can be inferred about their physiology.

Additional candidate divisions. Several additional candidate divisions have been identified based on environmental sequences alone, shown as outlined wedges in Fig. 1. These divisions comprise two or more sequences over 500 nt in length that were obtained mostly from independent studies, or at least from independent PCR events. An expanded view detailing representatives of each candidate division is available at <http://crab2.berkeley.edu/pacelab/176.htm>. The candidate divisions are identified according to the original source or clone names of the sequences that define the clade. Divisions designated OP were originally identified in an analysis of a Yellowstone hot spring, Obsidian Pool (21). Representatives of three of these divisions, OP5, -8, and -10, also have been encountered in a study of a hydrocarbon-contaminated aquifer at Wurtsmith Air Force Base in Michigan (9). The latter study also identified novel divisions WS1, now identified in a Siberian tundra soil (53), and WS6. Candidate division marine group A was originally identified and named based on partial sequences obtained from marine microbial communities in the Atlantic and Pacific oceans (13) and verified by full-length-sequence representatives of the group from similar marine samples (16). Abundance and depth profiles of marine group A sequences in the water column (16) suggest their global distribution in marine communities; no representatives of this candidate division outside of marine environments have yet been obtained (Table 1). Representatives of the termite group I candidate division originally were identified as a closely related clade of sequences from the termite gut (33) but now also have been identified in a contaminated aquifer (9). Candidate division OS-K was identified in a study of a Yellowstone hot spring, Octopus Spring (49) and bolstered by additional representative sequences from studies of a hydrothermal vent (32) and marine sediment (7). Candidate divisions TM6 and TM7 are named after sequences obtained in an environmental study of a peat bog (38), and other partial-length-sequence representatives of these candidate divisions were subsequently identified from activated sludges (4, 15) and soil (5).

CONCLUSION

Phylogenetic trees based on rRNA sequences show that bacterial diversity is represented by natural relatedness groups, the phylogenetic divisions (51). About 36 such divisions are currently identifiable. The final extent of division-level diversity in the bacterial domain is still unknown but clearly will be more than 40 divisions. Culture-independent studies have resulted in multiple hits on the majority of described divisions in different habitat types (Table 1), suggesting that the final number of divisions will be within the same order of magnitude as the present estimate.

The molecular analyses of environmental DNA have revealed substantial phylogenetic diversity with little or no representation among organisms previously studied. Because of their abundance and wide distribution, some of the organisms represented by the sequences likely contribute significantly to the global chemical cycles. Descriptions of newly identified, but apparently important, bacterial divisions such as the *Acidobacterium* and *Ferricimicrobia*, are presently confounded by too few cultivated representatives and only rudimentary descriptions of the strains. Cultivation efforts need to be directed at new representatives of the diverse groups for further study. Continued work to sequence the 16S rDNAs of all deposited type cultures (>500 sequenced to date [14]) may also result in

Verrucomicrobia division

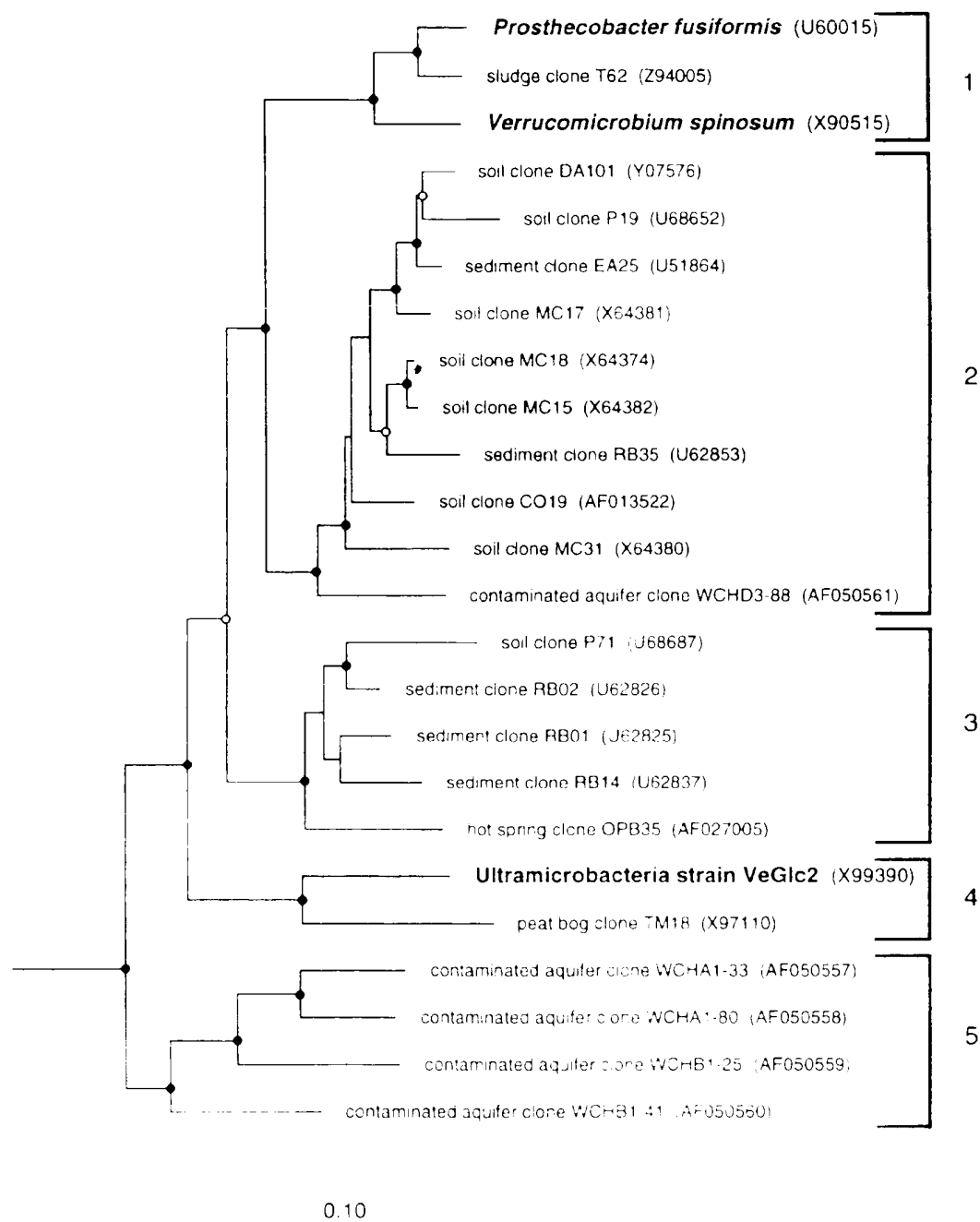


FIG. 4. Phylogenetic dendrogram of the *Verrucomicrobia* division. Names of cultivated organisms are in italics. The habitat source of each environmental sequence is indicated before the clone name. GenBank accession numbers are listed parenthetically. Systems (see the text) are indicated by brackets at the right of the tree. Tree construction and support for branch points was as described for Fig. 1 and 3, respectively. The scale bar indicates 0.1 change per nucleotide.

Detection of additional cultivated representatives of newly described divisions. It is a challenge to microbial biologists to determine the physiological diversity and environmental roles of these recently articulated divisions of *Bacteria*.

The phylogenetic differences between the bacterial divisions probably are reflected in substantial physiological differences. Some properties, the general properties of *Bacteria*, are expected to be distributed among all the divisions. Division-specific

Green non-sulfur division

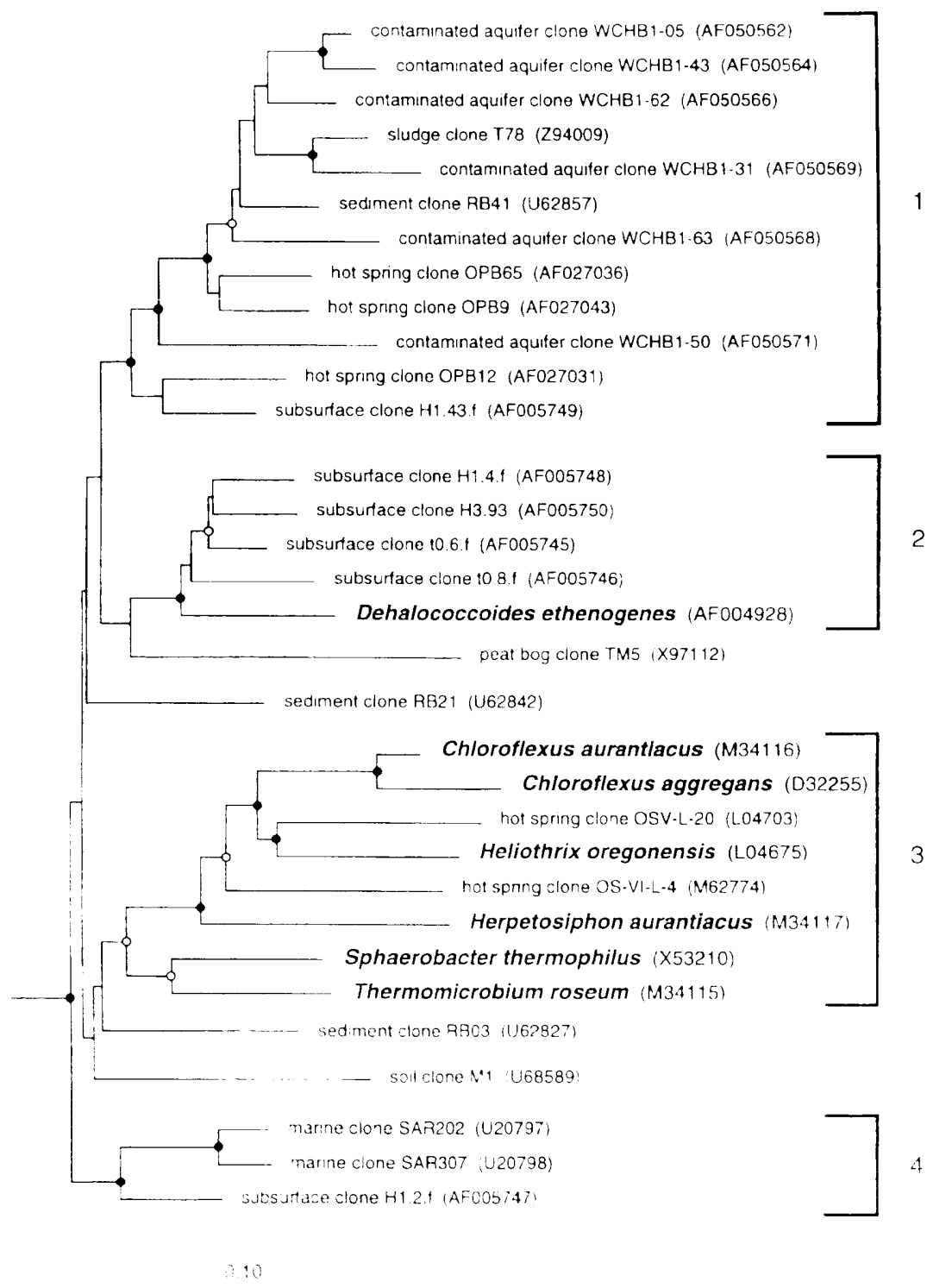


FIG. 5. Phylogenetic dendrogram of the Green non-sulfur division. Names of cultivated organisms are shown to the right. The highest source of similarity for each clone is indicated before the clone name (see Benoit et al., 1997, for details on related sequences). Subdivisions along the tree are indicated by brackets at the right. The tree construction and support for branching nodes are described for Fig. 1 and 3, respectively. The scale bar indicates 0.1 change per nucleotide.

OP11 division

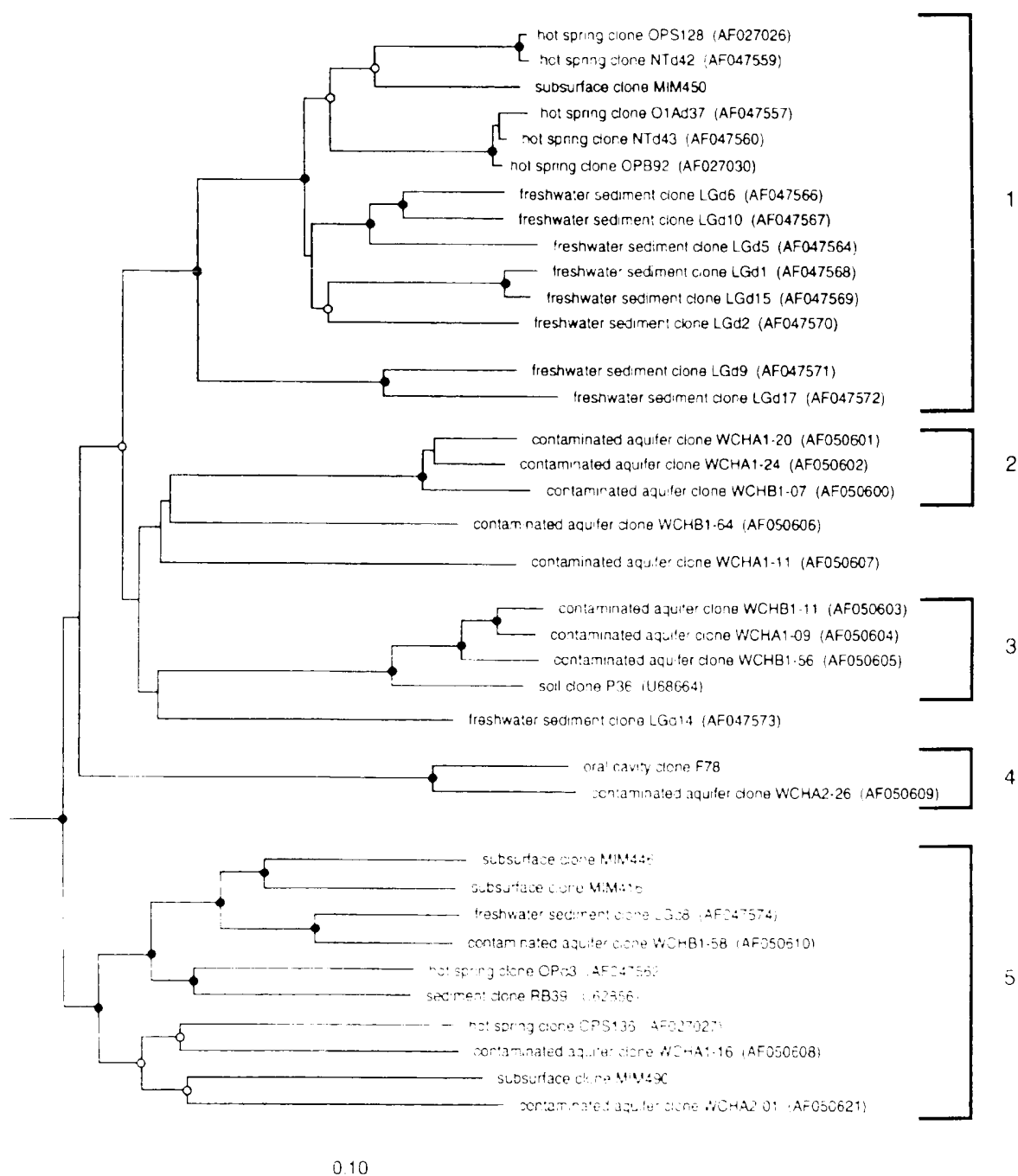


FIG. 1. Phylogenetic dendrogram of the OP11 division. The habitat source of each clone is indicated before the clone name. GenBank accession numbers are listed parenthetically. Subdivisions (see the text) are indicated by the numbers 1-5 on the right. Tree construction and support for each point was as described for Fig. 2 and 3 respectively. The four MIM clones and P36 clone were sequenced by the author; all other data were available to us by Philippe Durr and John and Joyce Johnson. Scale bar indicates 0.1 change per nucleotide.

novelties are known as well, for instance, endospore formation by the low-G+C gram-positive bacteria or axial filaments (endoflagella) in the spirochetes. Some biochemical properties evidently have transferred laterally among the divisions. For example, the two types of photosynthetic complexes, photosystem I (PSI) and PSII, are each distributed sporadically among the divisions, consistent with lateral transfer (3). Lateral transfer may also have resulted in combinatorial novelty among the divisions; PSI and PSII, for instance, apparently came together in the cyanobacteria to create oxygenic photosynthesis, with profound consequences to the biosphere (3). Many more such division-specific qualities and cooperations should become evident at the molecular level as comparative genomics gives us a sharper phylogenetic picture of bacterial diversity.

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REFERENCES

- Amann, R. L., W. Ludwig, and K. H. Schleifer. 1995. Phylogenetic identification and in situ detection of individual microbial cells without cultivation. *Microbiol. Rev.* 59:143-169.
- Barns, S. M., C. F. Delwiche, J. D. Palmer, and N. R. Pace. 1996. Perspectives on archaeal diversity, thermophily and monophyly from environmental rRNA sequences. *Proc. Natl. Acad. Sci. USA* 93:9188-9193.
- Blankenship, R. E. 1992. Origin and early evolution of photosynthesis. *Photosynth. Res.* 33:91-111.
- Bond, P. L., P. Hugenholtz, J. Keller, and L. L. Blackall. 1995. Bacterial community structures of phosphate-removing and non-phosphate-removing activated sludges from sequencing batch reactors. *Appl. Environ. Microbiol.* 61:1913-1916.
- Borneman, J., and E. W. Triplett. 1997. Molecular microbial diversity in soils from eastern Amazonia: evidence for intrasubstratum microorganisms and microbial population shifts associated with deforestation. *Appl. Environ. Microbiol.* 63:2647-2653.
- Chandler, D. P., J. K. Fredrickson, and E. J. Brockman. 1997. Effect of PCR template concentration on the composition and distribution of total community 16S rDNA clone libraries. *Mol. Ecol.* 6:475-482.
- Devereux, R., and G. W. Mundfrom. 1994. A phylogenetic tree of 16S rRNA sequences from sulfate-reducing bacteria in a sandy marine sediment. *Appl. Environ. Microbiol.* 60:3437-3439.
- Dewhirst, F. 1998. Personal communication.
- Dojka, M. A., P. Hugenholtz, S. K. Haack, and N. R. Pace. Microbial diversity in a hydrocarbon- and chlorinated-solvent-contaminated aquifer undergoing intrinsic bioremediation. *Appl. Environ. Microbiol.* in press.
- Durand, P. 1998. Personal communication.
- Ehrlich, S., D. Behrens, E. Lebedeva, W. Ludwig, and E. Bock. 1995. A new obligately chemolithoautotrophic, nitrate-oxidizing bacterium, *Nitrospira moscovensis* sp. nov., and its phylogenetic relationship. *Arch. Microbiol.* 164:16-23.
- Farrelly, V., F. A. Rainey, and E. Stackebrandt. 1995. Effect of genome size and *mcr* gene copy number on PCR amplification of 16S rRNA genes from a mixture of bacterial species. *Appl. Environ. Microbiol.* 61:2785-2801.
- Fuhrman, J. A., K. McCallum, and A. A. Davis. 1993. Phylogenetic diversity of subsurface marine microbial communities from the Atlantic and Pacific oceans. *Appl. Environ. Microbiol.* 59:1294-1302.
- Garrity, G. M., J. Tiedje, and D. Searles. 1995. Report on a workshop on the phylogeny of prokaryotes based upon sequence similarity of the small ribosomal subunit. Unpublished report.
- Godon, J.-J., E. Ziemstein, P. Dabert, F. Habouzit, and R. Moletta. 1997. Molecular microbial diversity of an anaerobic digester as determined by small-subunit rDNA sequence analysis. *Appl. Environ. Microbiol.* 63:2802-2813.
- Gordon, D. A., and S. J. Giovannoni. 1996. Detection of stratified microbial communities in the water column of *Hydrocarbon* in the Atlantic and Pacific oceans. *Appl. Environ. Microbiol.* 62:1171-1177.
- Hedlund, B. P., J. J. Gosink, and J. E. Staley. 1997. Genomic organization of a new locus of the bacterium *Thiomargarita* from a new species of the genus. *Arch. Microbiol.* 168:24-38.
- Hiraishi, A., S. Kishimoto, Y. Kosako, S. Wakao, and T. Tano. 1997. Phylogenetic position of the menaquinone-containing acidophilic chemolithoautotroph *Acidobacterium capsulatum*. *FEMS Microbiol. Lett.* 132:91-94.
- Hugenholtz, P., K. L. Hershberger, J. L. Flanagan, B. Kimmel, and N. R. Pace. 1997. Widespread distribution of a novel phylum-depth bacterial lineage in nature, abstr. N-23, p. 385. In Abstracts of the 97th General Meeting of the American Society for Microbiology 1997. American Society for Microbiology, Washington, D.C.
- Hugenholtz, P., and N. R. Pace. 1996. Identifying microbial diversity in the natural environment: a molecular phylogenetic approach. *Trends Biotechnol.* 14:190-197.
- Hugenholtz, P., C. Pitulle, K. L. Hershberger, and N. R. Pace. 1998. Novel division level bacterial diversity in a Yellowstone hot spring. *J. Bacteriol.* 180:366-376.
- Janssen, P. H., A. Schuhmann, E. Morschel, and F. A. Rainey. 1997. Novel anaerobic ultramicrobacteria belonging to the *Verrucomicrobiales* lineage of bacterial descent isolated by dilution culture from anoxic rice paddy soil. *Appl. Environ. Microbiol.* 63:1352-1358.
- Kopczynski, E. D., M. M. Bateson, and D. M. Ward. 1994. Recognition of chimeric small-subunit ribosomal DNAs composed of genes from uncultivated microorganisms. *Appl. Environ. Microbiol.* 60:746-748.
- Kuske, C. R., S. M. Barns, and J. D. Busch. 1997. Diverse uncultivated bacterial groups from soils of the arid and southwestern United States that are present in many geographic regions. *Appl. Environ. Microbiol.* 63:3614-3621.
- Lee, S.-Y., J. Bollinger, D. Bezdecik, and A. Ogram. 1996. Estimation of the abundance of an uncultured soil bacterial strain by a competitive quantitative PCR method. *Appl. Environ. Microbiol.* 62:3787-3793.
- Liesack, W., F. Bak, J. U. Kreft, and E. Stackebrandt. 1994. *Holophaga foetida* gen. nov., sp. nov., a new homoacetogenic bacterium degrading methylated aromatic compounds. *Arch. Microbiol.* 162:85-90.
- Liesack, W., R. Soller, T. Stewart, H. Haas, S. Giovannoni, and E. Stackebrandt. 1992. The influence of tachytelically (rapidly) evolving sequences on the topology of phylogenetic trees—infrafamily relationships and the phylogenetic position of Planctomycetaceae as revealed by comparative analysis of 16S ribosomal RNA sequences. *Syst. Appl. Microbiol.* 15:357-362.
- Loneragan, D. J., H. L. Jenter, J. D. Coates, E. J. P. Phillips, T. Schmidt, and D. R. Lovley. 1996. Phylogenetic analysis of dissimilatory Fe(III)-reducing bacteria. *J. Bacteriol.* 178:2402-2408.
- Ludwig, W., S. H. Bauer, M. Bauer, I. Held, G. Kirchhof, R. Schulze, I. Huber, S. Spring, A. Hartmann, and K.-H. Schleifer. 1997. Detection and *in situ* identification of representatives of a widely distributed new bacterial phylum. *FEMS Microbiol. Lett.* 153:181-190.
- Maidak, B. L., G. J. Olsen, N. Larsen, R. Overbeek, M. J. McCaughey, and C. R. Woese. 1997. The RDP (Ribosomal Database Project). *Nucleic Acids Res.* 25:109-111.
- Maymo-Gatell, X., Y.-L. Chien, J. Gossett, and S. Zinder. 1997. Isolation of a bacterium that reductively dechlorinates tetrachloroethene to ethene. *Science* 276:1568-1571.
- Moyer, C. L., E. C. Dobbs, and D. M. Karl. 1995. Phylogenetic diversity of the bacterial community from a microbial mat at an active hydrothermal vent system, Loihi Seamount, Hawaii. *Appl. Environ. Microbiol.* 61:1555-1562.
- Ohkuma, M., and T. Kudo. 1996. Phylogenetic diversity of the intestinal bacterial community in the territe *Rattus norvegicus*. *Appl. Environ. Microbiol.* 62:451-458.
- Pace, N. R. 1997. A molecular view of microbial diversity and the biosphere. *Science* 276:73-74.
- Pace, N. R., D. A. Stahl, D. J. Lane, and G. J. Olsen. 1986. The analysis of natural microbial communities by ribosomal RNA sequences. *Adv. Microbiol. Ecol.* 9:1-55.
- Reysenbach, A. L., L. J. Giver, G. S. Wickham, and N. R. Pace. 1992. Differential amplification of rRNA genes by polymerase chain reaction. *Appl. Environ. Microbiol.* 58:3417-3418.
- Reysenbach, A. L., G. S. Wickham, and N. R. Pace. 1994. Phylogenetic analysis of the hyperthermophilic, anaerobic community in Octopus Spring, Yellowstone National Park. *Appl. Environ. Microbiol.* 60:2113-2119.
- Rheims, H., F. A. Rainey, and E. Stackebrandt. 1996. A molecular approach to search for diversity among bacteria in the environment. *J. Ind. Microbiol.* 17:159-169.
- Schlegel, H. G., and H. W. Jannasch. 1997. Prokaryotes and their habitats, p. 3-135. In *A Practical Handbook of Microbiology*, W. H. H. H. Schleifer, ed. Chapman & Hall, New York, N.Y.
- Schmidt, T. M., F. E. DeLong, and N. R. Pace. 1994. Analysis of a marine picoplankton community by 16S rRNA sequencing and sequencing. *J. Bacteriol.* 173:4371-4375.
- Stackebrandt, E., R. G. E. Murray, and H. G. Truper. 1988. *Photobacterium* class (new class) in the phylum *Proteobacteria* that includes the purple bacteria and the green sulfur bacteria. *Arch. Microbiol.* 150:221-228.
- Stackebrandt, E., F. A. Rainey, and N. L. Ward-Rainey. 1997. Proposal for a new phylum, the *Proteobacteria*, in the kingdom *Bacteria*. *J. Syst. Bacteriol.* 47:133-134.
- Strunk, O., O. Gross, B. Reichen, M. May, S. Hermann, S. Struckmann, B.

- Nonhoff, M., Lenke, A., Vilbig, T., Ludwig, A., Bode, K., H. Schleifer, and W. Ludwig. 1998. ARB: a software environment for sequence data. Submitted for publication.
44. Suzuki, M. T., and S. J. Giovannoni. 1996. Bias caused by template annealing in the amplification of mixtures of 16S rRNA genes by PCR. *Appl. Environ. Microbiol.* **62**:625-630.
45. Tanner, M., B. M. Goebel, M. A. Dojka, and N. R. Pace. 1998. Specific rDNA sequences from diverse environmental settings correlate with experimental contaminants. Submitted for publication.
46. Ward, D. M., R. Weller, and M. M. Bateson. 1990. 16S rRNA sequences reveal numerous uncultured microorganisms in a natural community. *Nature* **345**:63-65.
47. Ward-Rainey, N., F. A. Rainey, H. Schlesner, and E. Stackebrandt. 1995. Assignment of hitherto unidentified 16S rDNA species to a main line of descent within the domain *Bacteria*. *Microbiology* **141**:3247-3250.
48. Weisburg, W. G., S. M. Barns, D. A. Pelletier, and D. J. Lane. 1991. 16S ribosomal DNA amplification for phylogenetic study. *J. Bacteriol.* **173**:697-703.
49. Weller, R., M. M. Bateson, B. K. Heimbuch, E. D. Kocczynski, and D. M. Ward. 1992. Uncultivated eubacteria, *Chloroflexus*-like inhabitants, and spirochete-like inhabitants of a hot spring microbial mat. *Appl. Environ. Microbiol.* **58**:3964-3969.
50. Weller, R., J. Weller, and D. M. Ward. 1991. 16S rRNA sequences of uncultivated hot spring cyanobacterial mat inhabitants retrieved as randomly primed cDNA. *Appl. Environ. Microbiol.* **57**:1146-1151.
51. Woese, C. R. 1987. Bacterial evolution. *Microbiol. Rev.* **51**:221-271.
52. Woese, C. R., S. Winker, and R. R. Gutell. 1990. Architecture of ribosomal RNA: constraints on the sequence of tetra-loops. *Proc. Natl. Acad. Sci. USA* **87**:8467-8471.
53. Zhou, J., M. E. Davey, J. B. Figueras, E. Rivkina, D. Gilichinsky, and J. M. Tiedje. 1997. Phylogenetic diversity of a bacterial community determined from Siberian tundra soil DNA. *Microbiology* **143**:3913-3919.